

Translation Regulation with Artificial Introns

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SCHOOL OF BIOLOGICAL AND HEALTH SYSTEMS ENGINEERING

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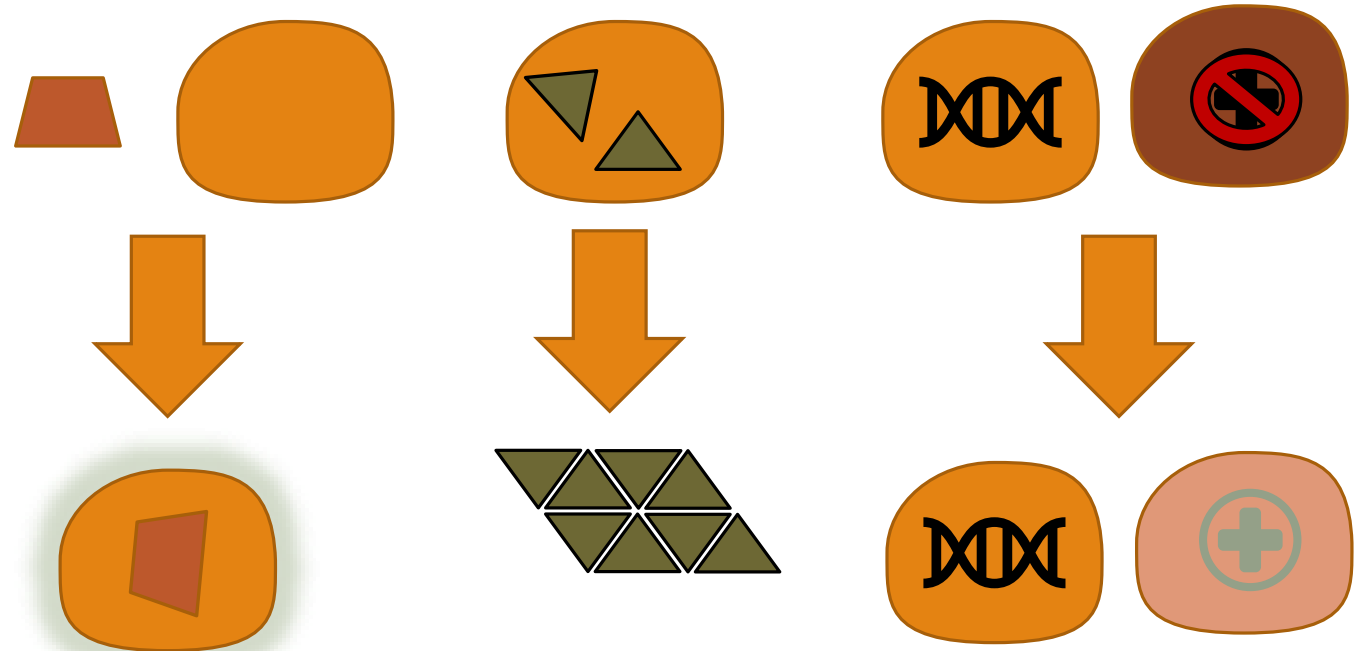


What is Synthetic Biology?

Design and control organisms

- Biosensors
- Compound and material production
- Therapies

Cellular Factories



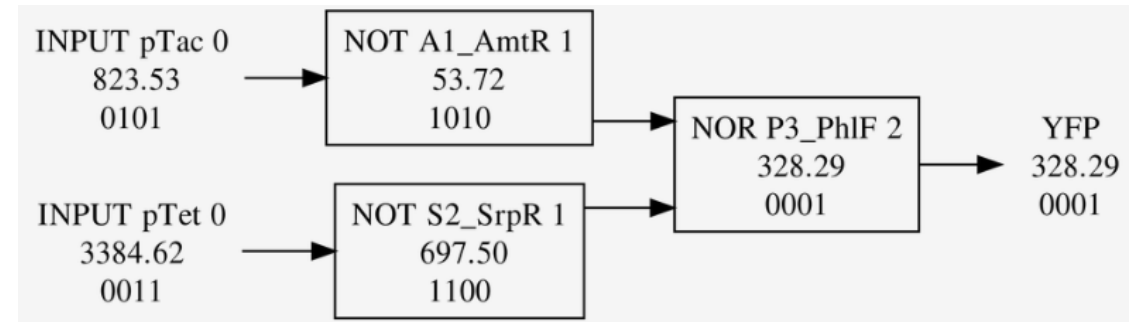
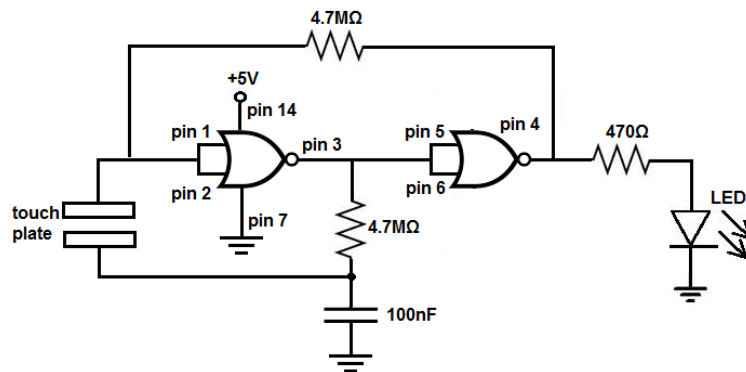
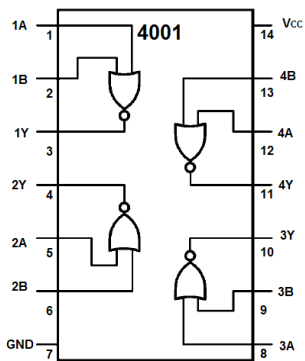


Synthetic Biology & Electrical Engineering

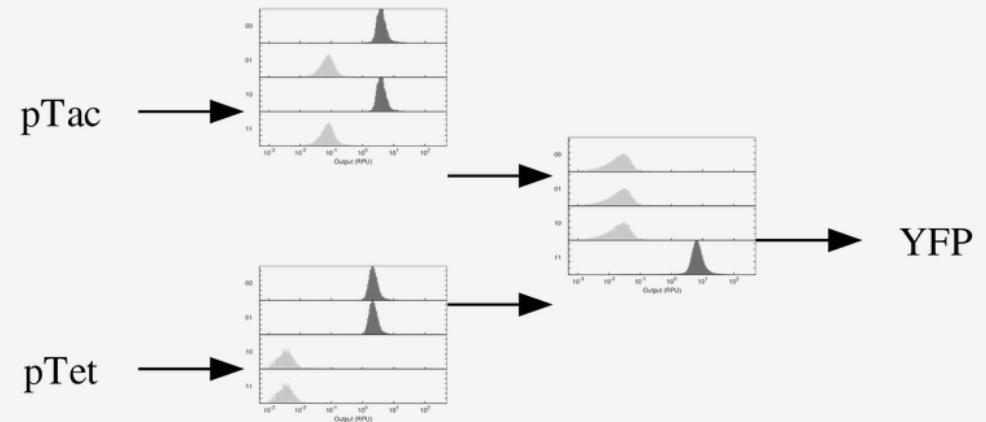
Integrated circuits are essential for the development of modern computing

Combination of genetic components to form complex gene circuits provide new solutions

Program biological systems to make decisions based on their environment and stimulus



predicted gate RPU's



(Cello)

(How to Build a Touch On-Off Circuit with a 4001 NOR Gate Chip)



Synthetic Biology Advantages

Ability to utilize chemistry and create compounds

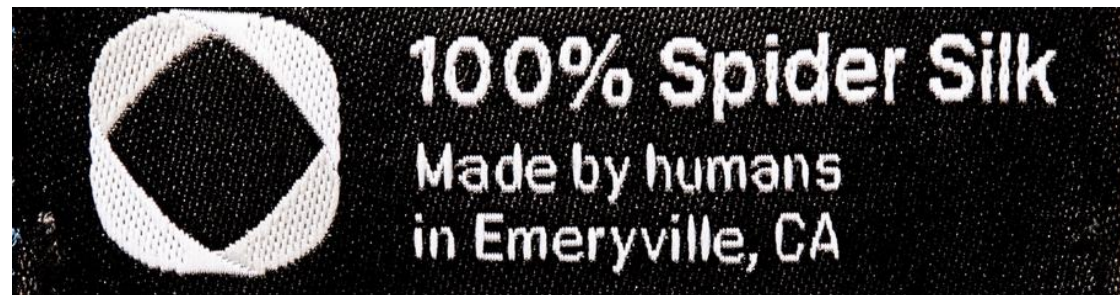
Directly interact with chemical stimuli

Ability to multiply and allow for scaling



Designing The Future of Gene and Cell-Based Therapies

Bolt Threads is harnessing proteins found in nature to create fibers and fabrics with both practical and revolutionary uses.



Ginkgo Bioworks is the organism company. We design custom microbes for customers across multiple markets. We build our foundries to scale the process of organism engineering using software and hardware automation. Organism engineers at Ginkgo learn from nature to develop new organisms that replace technology with biology.

(Amyris) (Bolt Threads) (“The Organism Company”) (“Senti Biosciences”)

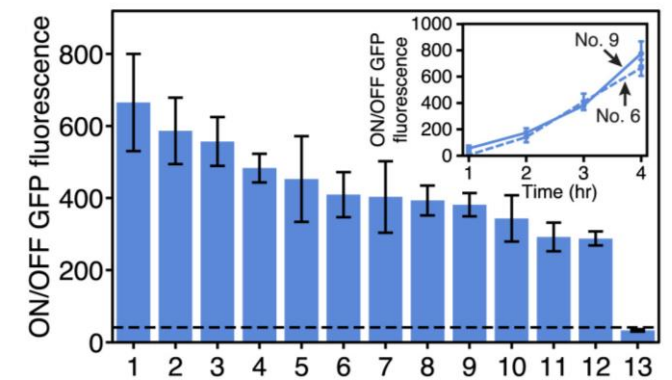
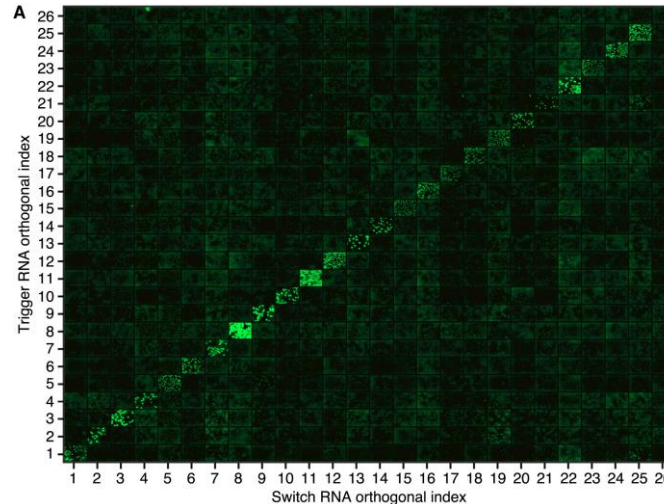
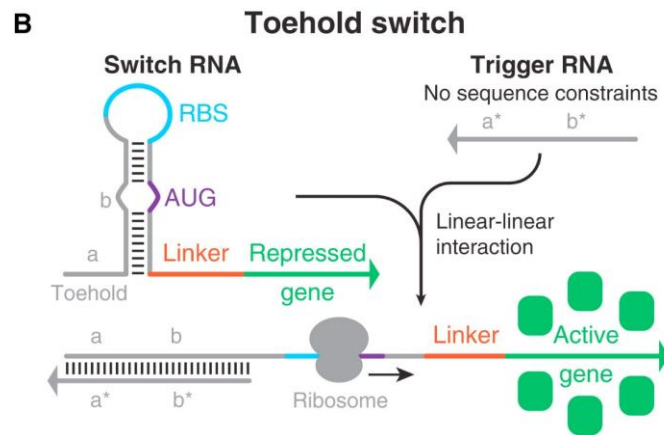
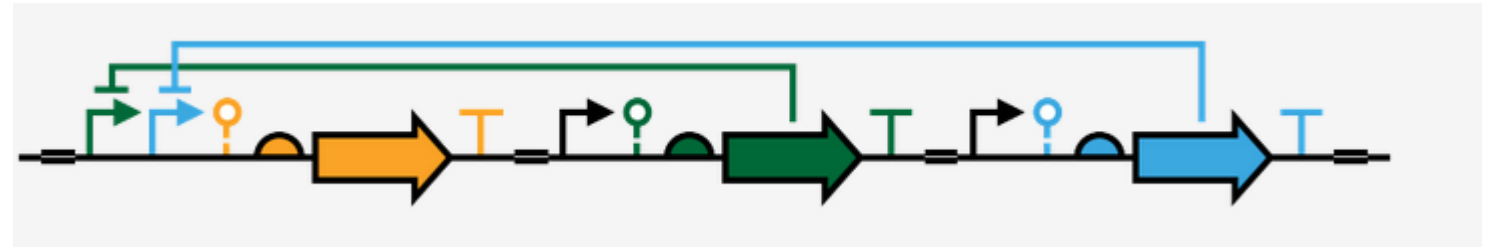


Project Inspiration: Gene Regulation Parts

Gene regulation parts are essential for creating genetic circuits

- Modular
- High Specificity
- High Dynamic Range

Toehold switch



(Green et al., 2014) (Cello)



Artificial Intron Demonstrates Directional Specific Splicing

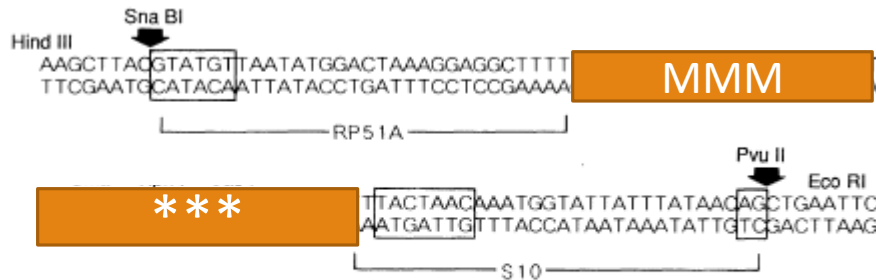
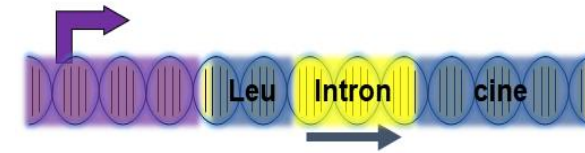
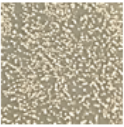

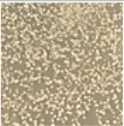



Fig. 1. Structure of the artificial intron. The 5' end fragment and the 3' end fragment were chemically synthesized, and the polylinker fragment (from Pst I to Sac I) was obtained from plasmid pUC18. The consensus sequences are shown in boxes. This intron was cloned in the Hind III–Eco RI site of pUC18 to generate pUC-*AI*. The 5' end fragment of the yeast *RP51A* intron and the 3' end fragment of the *S10* intron are indicated.

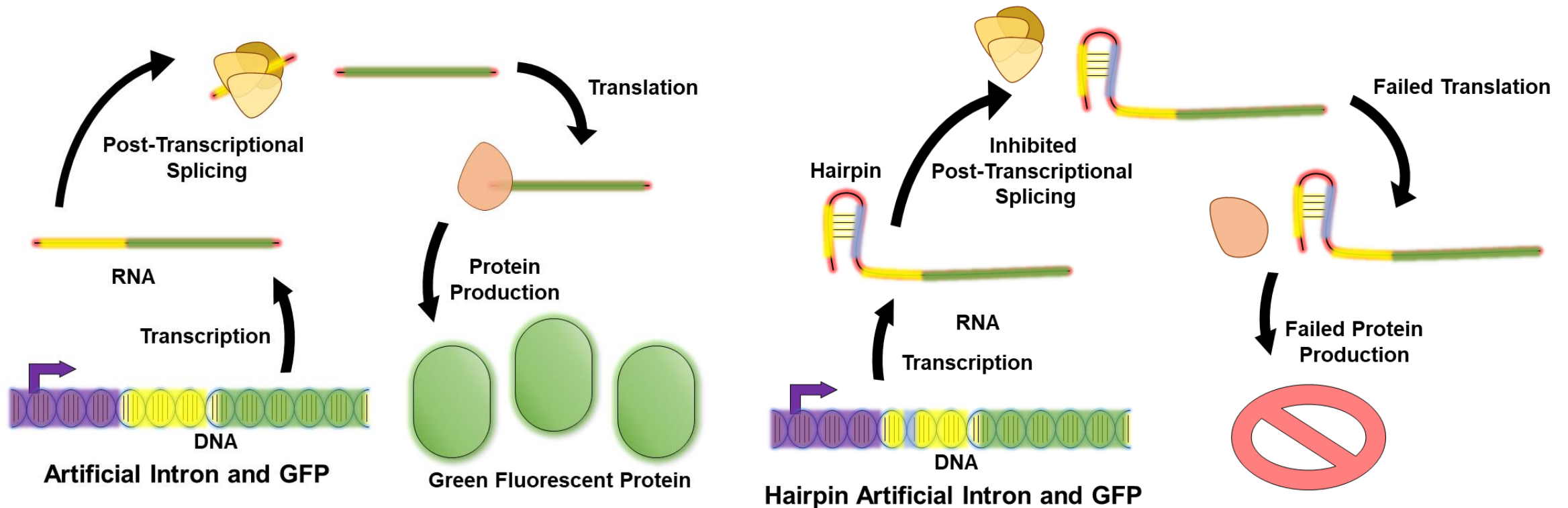
(Yoshimatsu and Nagawa, 1989)



 Intron 1 (Forward) (Leucine Production = Successful Splicing)	 Intron 2 (Backward) (No Leucine Production = Unsuccessful Splicing)
 Control +Leu2 (Leucine Production = Survival)	 Control -Leu2 (No Leucine Production = No Survival)

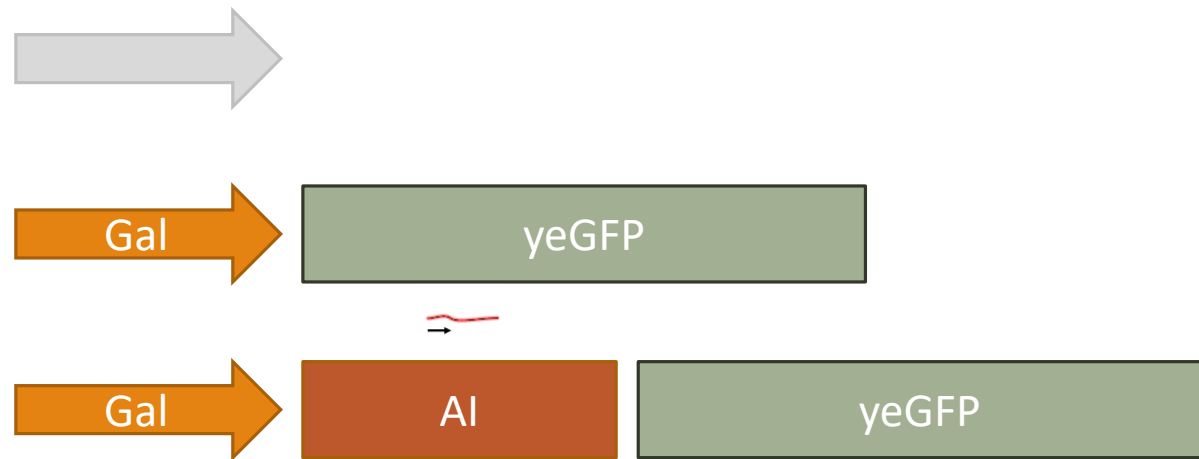
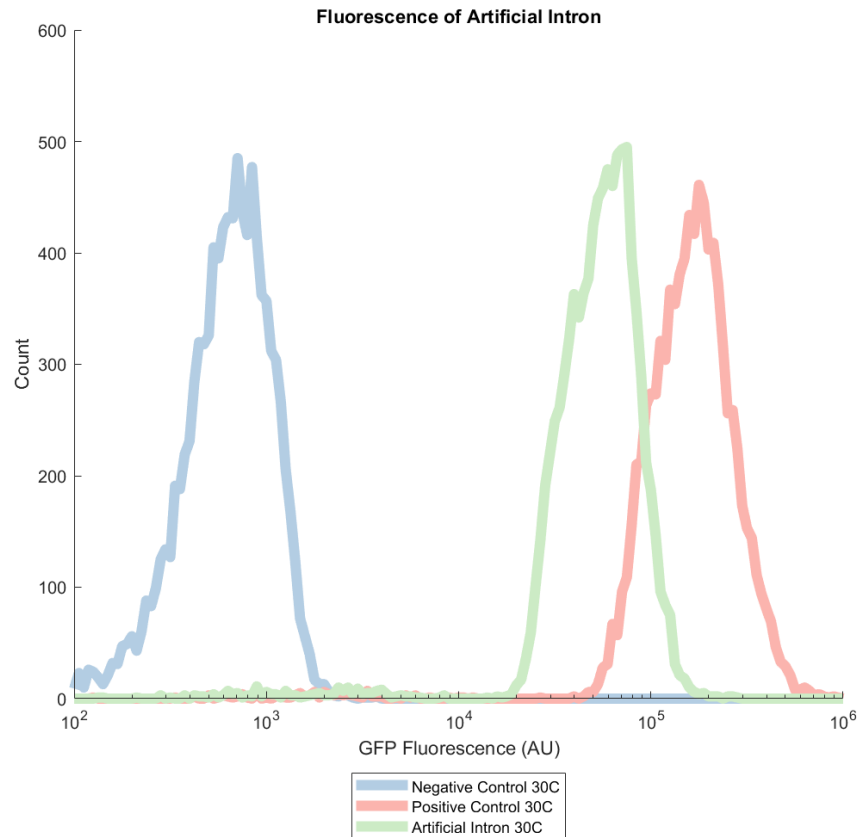


Hairpins Designed to Inhibit Splice Site Recognition



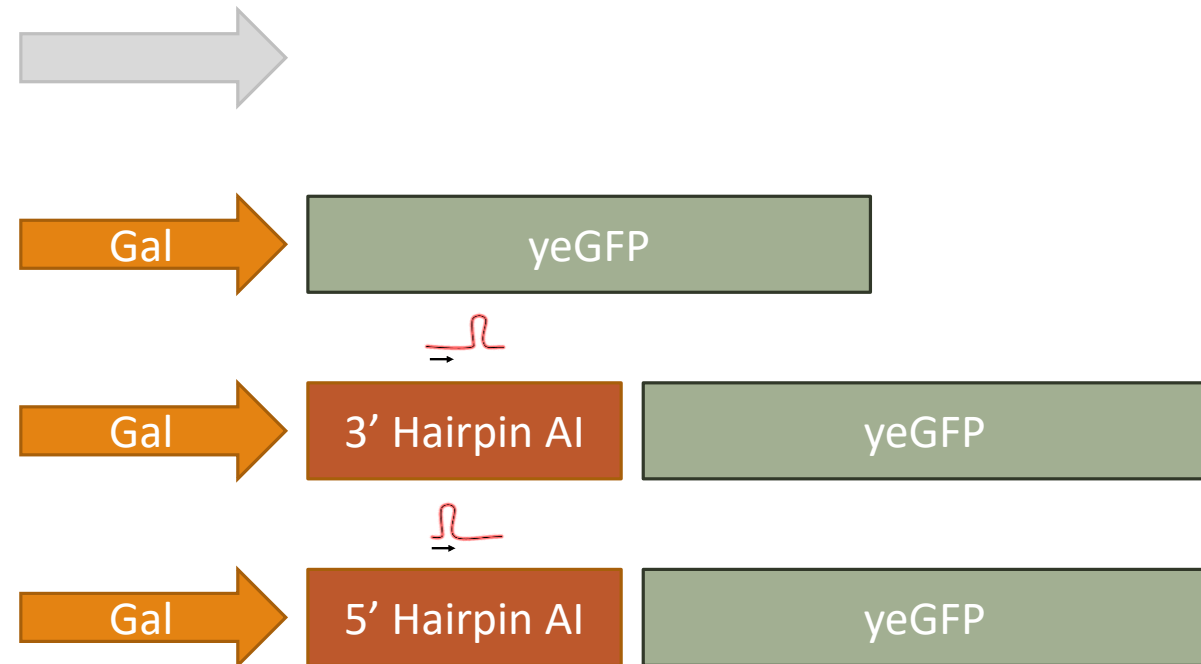
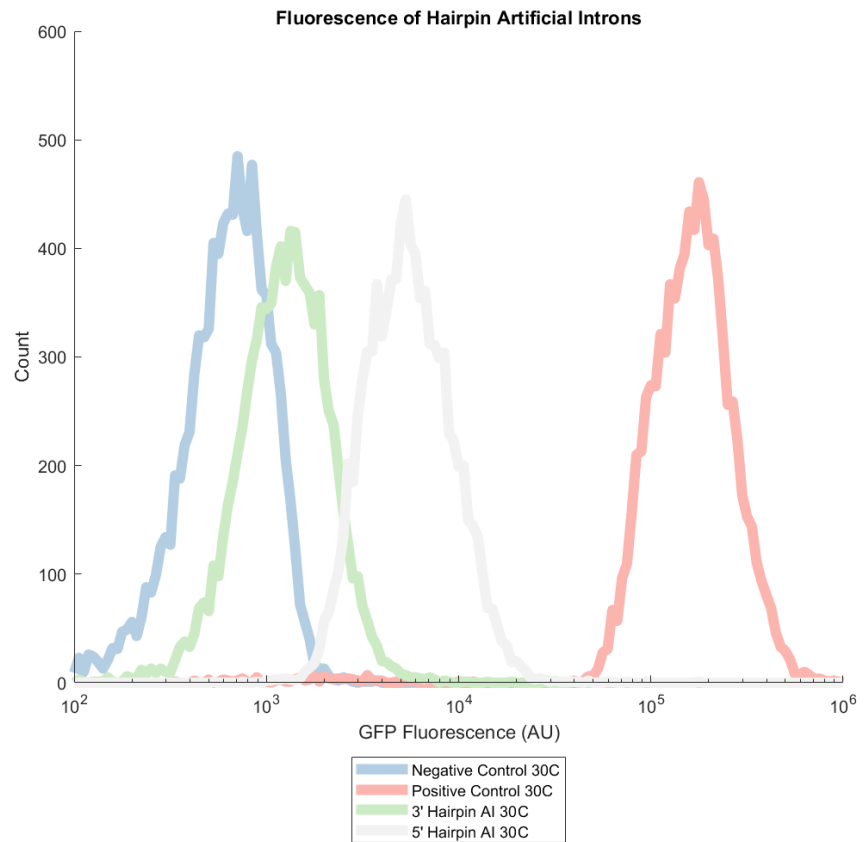


Intron Splicing Inhibition with Hairpins



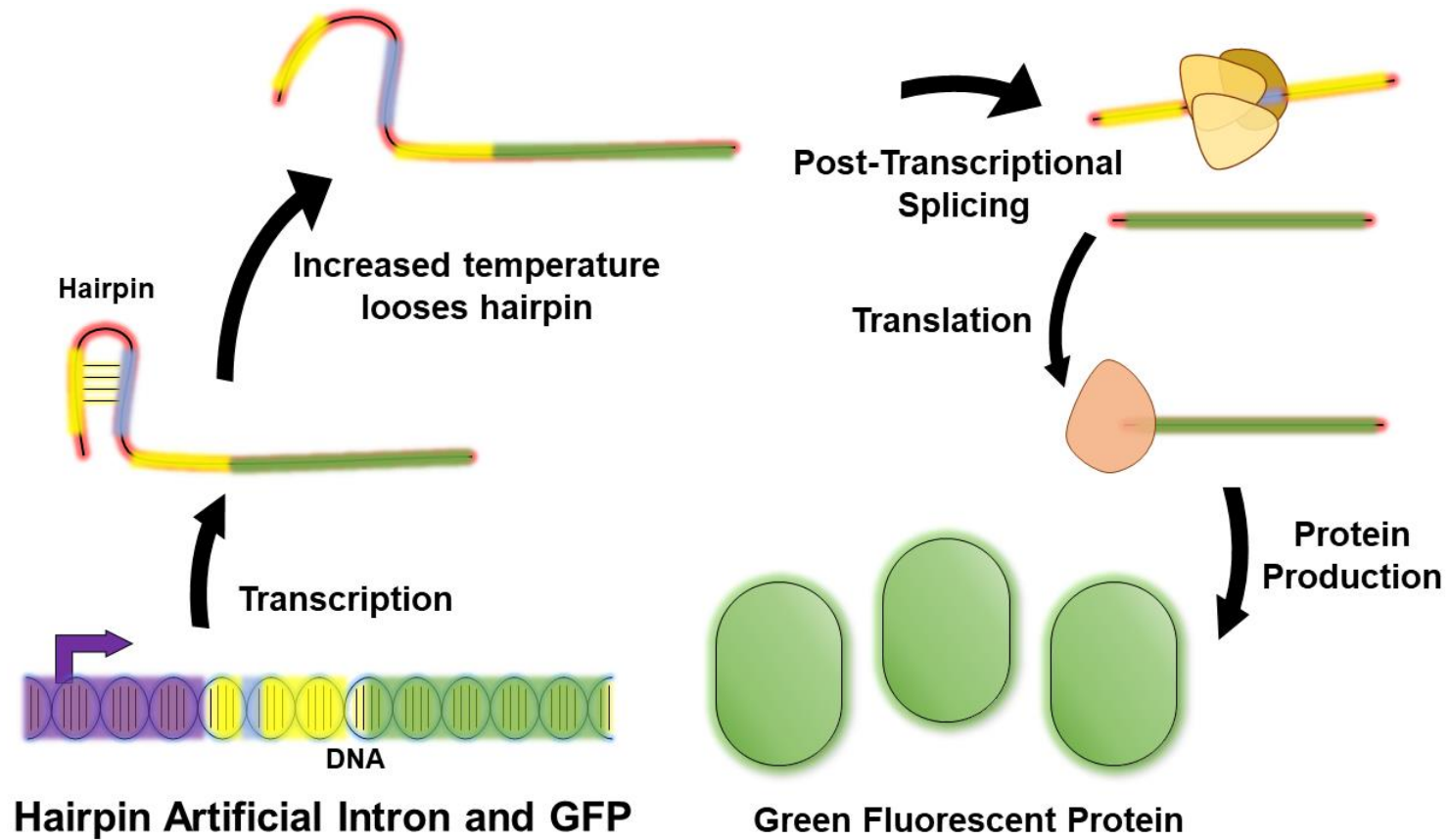


Intron Splicing Inhibition with Hairpins



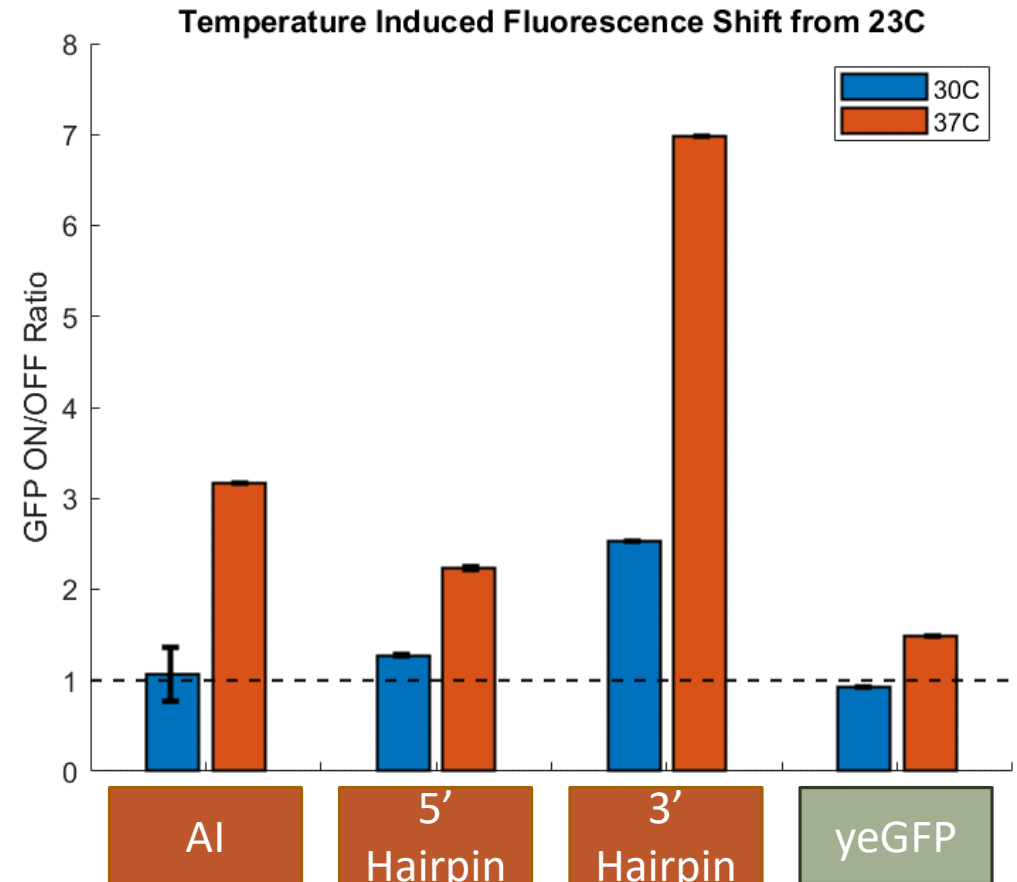
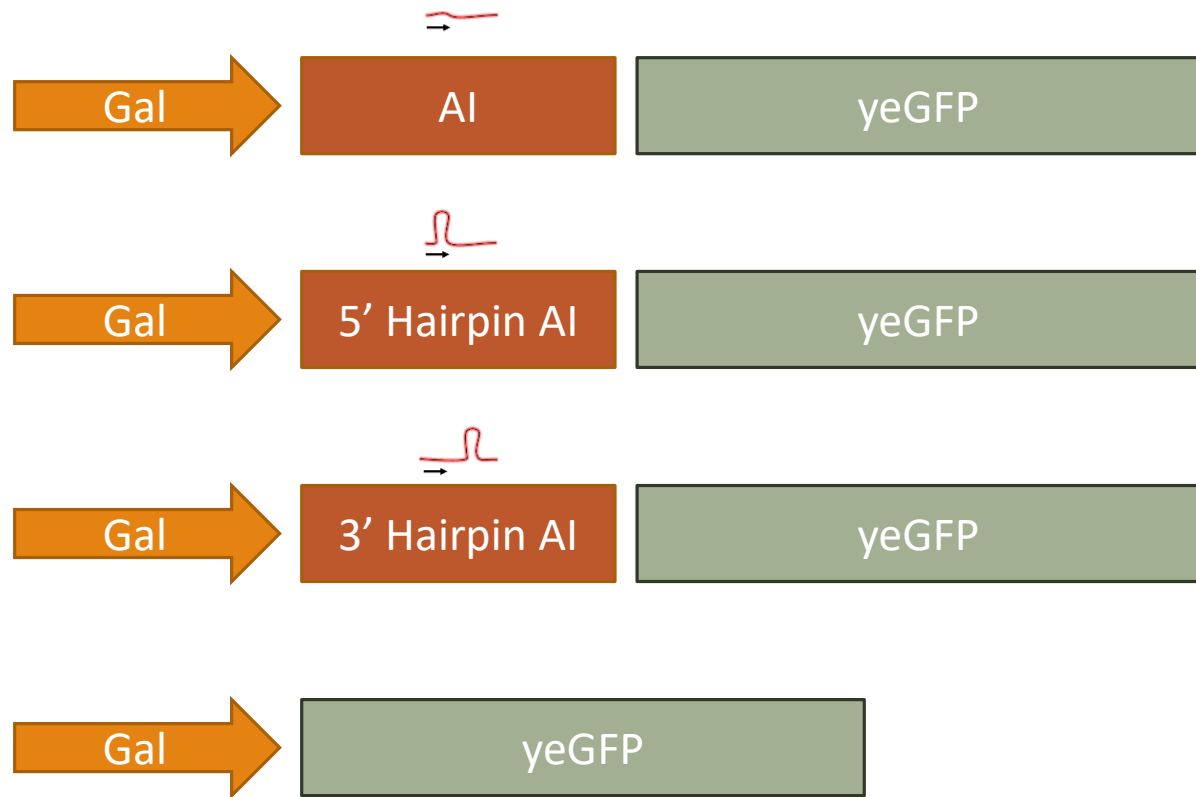


Temperature Sensitivity of Intron Splicing Inhibition





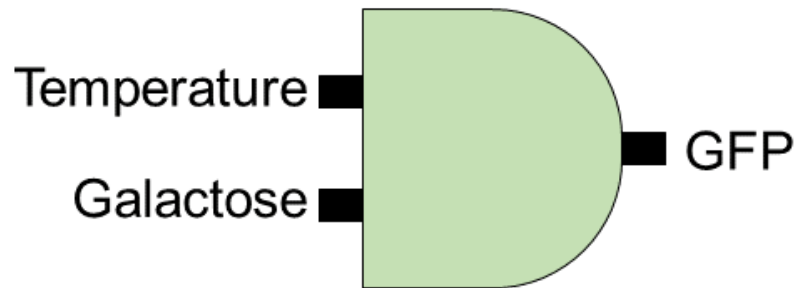
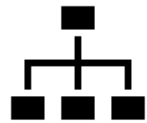
Temperature Sensitivity of Intron Splicing Inhibition



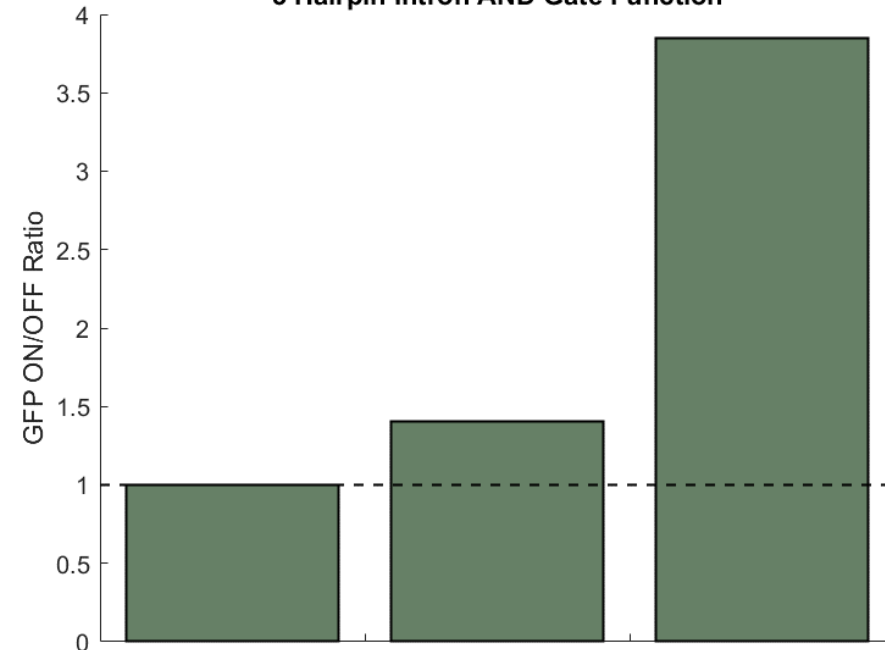
*AI = Artificial Intron



Logic AND Gate Functionality



3 Hairpin Intron AND Gate Function



Input

Temperature

-

-

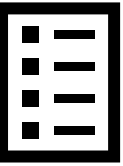
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Galactose

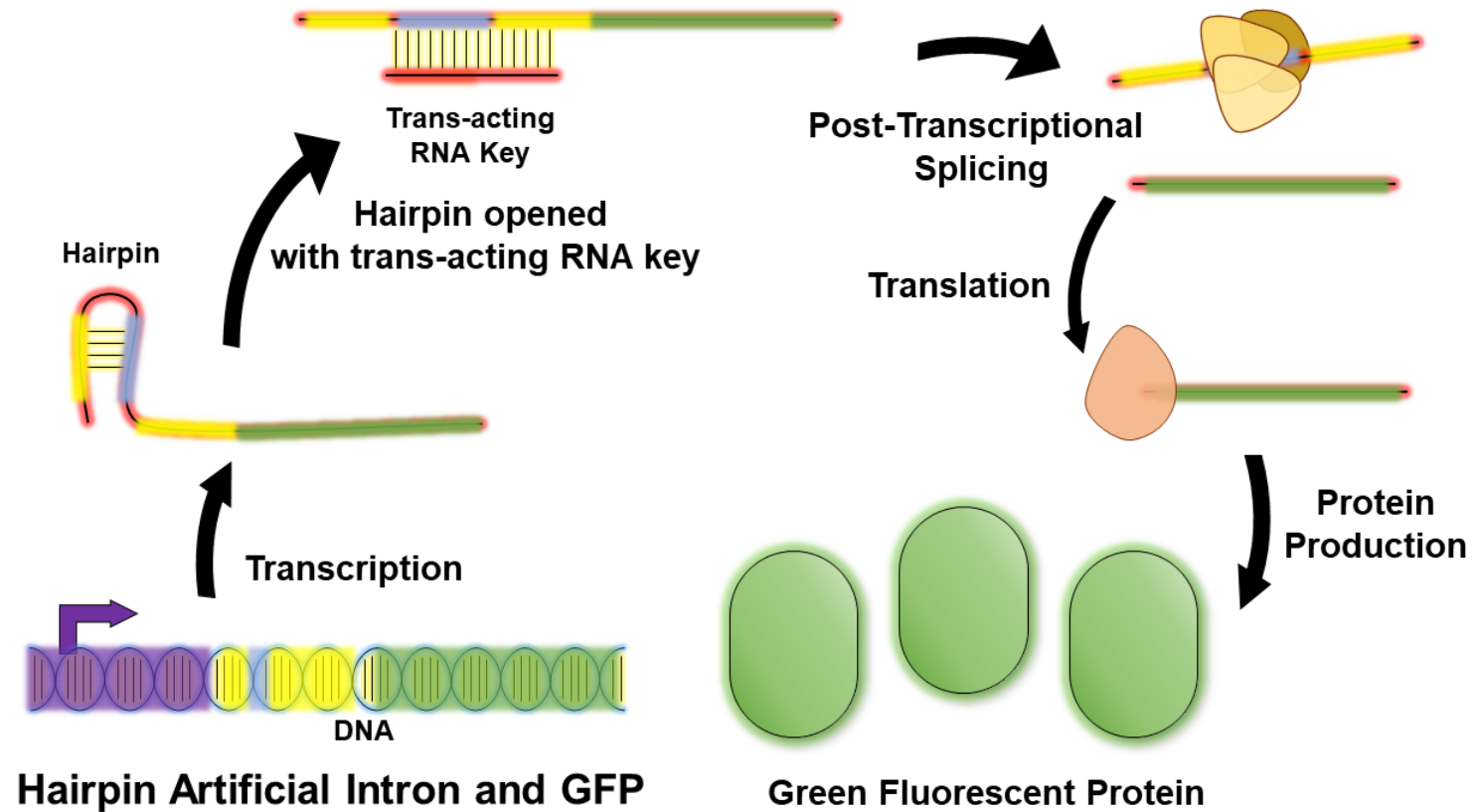
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Future Steps: *trans*-acting RNAs to control intron splicing



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xiao wang lab
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